

Physico-chemical and Nutritional Characterization of Dried Mangoes Consumed in Chad for Better Post- Harvest Valorization

ABSTRACT

Aim: To determine chemical and nutritional characterization of dried mangoes consumed in Chad for better post-harvest valorization.

UNDER PEER REVIEW

Place and Duration of Study: Food Sciences and Nutrition Research Laboratory (LARSAN), Faculty of Human Health Sciences, University of N'Djamena BP1117, Chad; between February 2022 and April 2023.

Methodology: Dried mangoes were collected from five localities (Doba, Bebedjia, Koumra, Moundou, and Bongor) were labeled DbA, Bja, Mdou, Kmr, and Bgr, and then stored in a refrigerated cooler for physico-chemical analyses using standard methods was used to determine chemical composition of various samples.

Results: These analyses showed that water content dried mangoes was not significantly influenced ($p = 0.0014$) by the production area. Protein, carbohydrate, lipid, and energy contents per 100 g ranged from 3.89 to 3.93 g, 94.91 to 104.72 g, 1.35 to 2.07 g, and 360.48 to 381.36 kcal/100 g, respectively. The reducing sugars content was high (77.73 g/100g) in Doba samples, compare to that from Moundou (76.51 g/100g). Starch and amylopectin contents were not significantly affected by production area (110.32 to 112.07 g/100g and 94.28 to 97.20 g/100 g, respectively). Calcium (Ca), sodium (Na), potassium (K), magnesium (Mg), and iron (Fe) were present in all samples with respective contents varying from 72.66 to 73.49 mg/100 g, 11.41 to 12.58 mg/100 g, 79.15 to 85.97, 68.78 to 70.59 mg/100 g, and 1.05 to 1.79 mg/100 g. Vitamin C and carotenoid contents varied, respectively, from 69.37 to 70.15 $\mu\text{g}/100\text{ g}$ and from 249.25 to 274.15 $\mu\text{g}/100\text{ g}$. Principal Component Analysis revealed that samples E1DOBA and E2BGA and E3Kmr and E4Mdr have the same physico-chemical characteristics.

Conclusion: Thus, these results demonstrate that respecting the drying and storage conditions for dried mango fruits, regardless the zone, it would enable the best possible preservation of nutritional properties while ensuring year-round availability.

Keywords: Physico-chemical properties; dried mangoes; production area; Chad.

1. INTRODUCTION

Chad is a country located in Central Africa with a suitable climate for the cultivation and development of fruits, vegetables, and citrus fruits. Among them, mango occupies an important place in production, marketing, and export [1], hence its appellation of "King of Fruits." More than 50 million metric tons are produced per year, and less than 4% of this production is exported [2,3]. In Chad, it represents the most important national fruit production and is therefore considered a jewel for the country's economic growth [4]. Average annual national mango production stands at 13,000 metric tons and is expected to double by 2020 [5]. Mangoes are produced, processed, and consumed in almost all parts of the country, including Bongor, Moundou, Doba, Bebedjia, and Koumra. Several dozen varieties from orchards in different localities have been recorded on the markets, with a preponderance of Amélie, Brooks, Kent, Keitt, Lippens, and Springfield varieties [6]. These are mainly sought after for their sweet flavors, soft textures, and rich nutrients [4].

Mangoes sold on market stalls in Chad are usually fresh, which poses storage problems, as they are no longer viable after 3 to 4 days, resulting in post-harvest losses that can amount

to over 40% of production [7]. This is all the more pronounced in Chad, where the average temperature is between 25 and 40°C, making it ideal for the growth of mesophilic pathogens [8].

To compensate and limit this loss, arboriculturists and merchants often use preservation methods using chemical agents such as formalin, citric acid, benzaldehyde, ash, etc., most of which are not without consequences for physico-chemical and hygienic qualities of the product [4]. What's more, adding value to this fruit by transforming it into a wide range of by-products such as juices, nectars, jams and syrups may cause problems such as low financial resources for equipment acquisition and lack of knowledge about the processes involved [7,9-10].

Thus, drying as a means of preservation appears to be an ideal solution, as it is inexpensive and easy to master [11]. It is also favored by the right climatic conditions (year-round sunshine and low humidity) and accounts for 80% of artisanal mango processing via solar drying and drying in airtight ovens. Although this solution may seem like a panacea, it is still in its infancy due to the low capacity of artisanal units, the lack of conservation/processing infrastructures and an unskilled workforce [10]. Failure to master these processing methods (drying) also affects several other parameters, such as organoleptic

properties like color, which is the main element influencing marketability [9]. It also affects physico-chemical properties, notably nutrient content, which can be lost during drying. These observations justify the importance of controlling drying conditions [12].

Mango plays a very important nutritional role, providing micronutrients required for the proper functioning of the human body and protecting against major chronic diseases such as cardiovascular, neurodegenerative and metabolic diseases, as well as cancers [13,14]. It's an excellent source of carbohydrates, β -carotene (provitamin A), vitamin C, polyphenols and fibre [15,8,16-23].

Consumption of half a portion of fruit contributes 100% of the recommended daily intake of vitamin A and up to 70% of that of vitamin C [11]. Mango pulp, made up of around 80% water and 20% dry matter, is recognized for its high nutritional value and medium energy intake (60 kcal per 100 g of pulp). The presence of provitamin A makes this fruit an important element in the fight against vitamin A deficiency and its corollaries, affecting more than 250 million children under 5 worldwide, with a high prevalence in Africa [24,25]. Alongside these vitamins and minerals, it also provides fiber and antioxidant compounds

with numerous biological properties (notably anti-diabetic) [26,27]. However, its composition depends on several intrinsic and extrinsic factors, such as variety, geographical area, climate, ripening stage, storage conditions, etc. [28,29]. (Martin and He, 2009; Tharanathan et al., 2006); and therefore can influence physico-chemical properties of some fruits such as mangoes.

Thus, the aim of the present study was to evaluate the impact of drying methods on physico-chemical properties of dried mangoes from five geographical areas in Chad.

2. MATERIALS AND METHODS

2.1 Study Framework and Period

The study was experimental and prospective. It took place in four Provinces of the Meridionale zone, namely the Province of *Logone Occidentale*, more precisely in Moundou, the Province of *Logone Orientale* in the localities of Doba and Bebedjia, the Province of *Mandoul* in the locality of Koumra and finally the Province of *Mayo Kebi Est* in the area of Bongor. These different provinces and localities were chosen for their high production, consumption and marketing of dried mangoes.

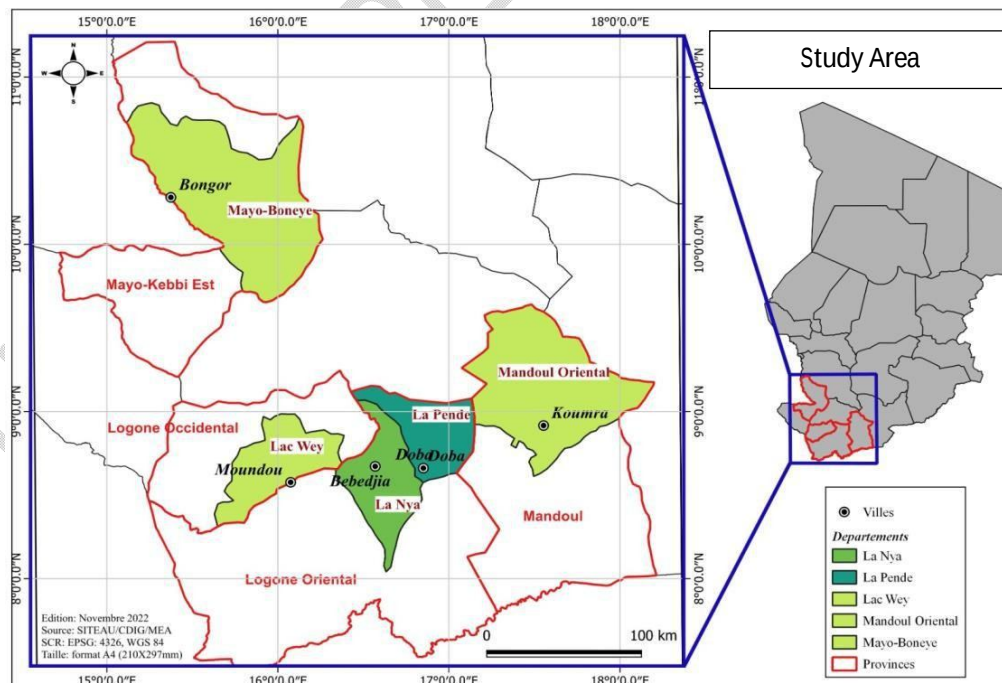


Fig.1. Study and sample collection area

2.2 Sampling

Once the various study areas had been identified, 100 samples of dried mangoes were collected from the various vendors in order to obtain an overall representation of the locality (20 samples per locality). After that, the samples were driven in a refrigerated cooler to the lab. After arriving at the laboratory, the dried mangoes were crushed, packaged in polystyrene plastic, and kept in a desiccator so they could be used in the various assays later on.

2.3 Determination of Chemical Composition of dried Mango Samples

Water content is determined according to the AACC method [30]. Thus, the vacuum cup is first cleaned, dried and weighed (M_0). After what, the cup containing the sample (5g) is weighed again (M_1) then placed in an oven at 105 °C for 3 hours. After this drying time, the cup is taken out of the oven, then cooled in a desiccator (P_2O_5) before being weighed (M_2) again. The results expressed represent the average of four tests and the water content (WC) is given by the following formula.

$$WC = (M_1 - M_2) / (M_1 - M_0) \quad (1)$$

M_0 : Mass of vacuum cup;

M_1 : Mass of sample and vacuum cup before drying;

M_2 : Mass of sample and vacuum cup after drying

Protein content was determined using the Kjeldahl method, which consists in converting organic nitrogen into ammoniacal nitrogen by mineralization with concentrated sulphuric acid, thus quantifying protein [30]. Formula 1 below was used to convert the rate of nitrogen release into protein:

$$\text{Protein content (\%)} = N (\text{rate of nitrogen release}) \times 6.25 (\text{conversion factor}) \quad (2)$$

Lipids were extracted with Soxhlet for 7 h using hexane as solvent. They were expressed as a percentage of dry matter according to [30]. The standard method described by AOAC (1990) was used to determine fiber content after digestion with 1.25% sulfuric acid for 30 min, followed by vacuum extraction and oven drying before weighing. The AOAC (1990) standard method was used to quantify total ash [30].

The AOAC (1990) standard method enabled us to quantify digestible carbohydrates in all samples by difference according to formula 3:

$$\text{Protein content (\%)} = 100 (\% \text{water} + \% \text{proteins} + \% \text{lipids} + \% \text{ash} + \% \text{total fiber}) \quad (3)$$

The energy values of the diets were obtained from the sum of the products of each major nutrient (carbohydrates, proteins, lipids) and its corresponding Atwater heat coefficient.

The Fisher and Stein 3,5-Dinitrosalicylic acid (DNS) method was used to quantify reducing sugars [31]. The lugol (iodine-iodide solution) method by Jarvis and Walker (1993) was used to determine starch content [32]. The colorimetric method of Chrastyl (1987) was used to quantify amylose. The amylopectin content was obtained by deduction between starch and amylose according to the formula below:

$$\text{Amylopectin (\%)} = 100 - \% \text{amylose} \quad (4)$$

Minerals such as calcium (Ca), magnesium (Mg), sodium (Na), potassium (K), zinc (Zn), (Fe) and copper (Cu) were determined on the ashes after prior digestion with HNO_3/HCl according to the standard protocol of AOAC (1990). Some were complexed before reading on a Perkin-Elmer Analyst 700 Atomic Absorption Spectrophotometer (AAS) in Norwalk, CT, USA, while others were read directly after digestion. Ca/Mg and Na/K mass ratios were also assessed.

2.4 Calorific Value, Recommended Daily Allowance (RDA)

The contribution of calorific value, protein and lipid content to the Recommended Daily Allowance (RDA), expressed as a percentage (%), was obtained from the following formula 5:

$$RDA (\%) = X/Y \times 100 \quad (5)$$

Where X represents the content or energy value per 100 g of this food and Y the recommended intake.

2.5 Determination of Carotenoid and Vitamin C Content

Using High Performance Liquid Chromatography (HPLC, AGILENT 1100), the β -carotene content of the lyophilisates of dried mango slices was ascertained. An Eppendorf tube containing a suspension in 10 ml of tetrahydrofuran (THF) solution was thoroughly shaken.

2.5 Statistical Analysis

The analysis was carried out using the ANOVA test (analysis of variance test) followed by Fisher post hoc test to compare means. The results were expressed as mean $\pm$ standard error of the mean. All these analysis were carried out by using Minitab 18.0 software. The value of probability $p < 0.05$ were considered as significant. A Principal Component Analysis was performed using XLSTAT version 2014 software to group the different sources of dried eats according to composition.

3. RESULTS AND DISCUSSION

3.1 Approximate Chemical Composition of Dried Mangoes

Table 1 below shows the approximate composition of dried mangoes from five different geographical areas.

3.2 Mineral Composition of Dried Mango Fruits from Five Locations in Chad

The Mineral composition of dried mangoes from five different geographical areas is shown in Table 2.

It appears that all samples contains minerals. No significant differences was observed between mineral concentrations and the five production area. However all the samples have high amount of potassium, magnesium and calcium compare to that of iron and sodium.

3.3 Vitamin Composition of Dried Mangoes from Five LOCATIONS in Chad

The vitamin composition of dried mangoes from five different geographical areas is shown in Table 3.

No significant difference ($P > 0.05$) was observed in Vitamin composition of dried mangoes from the five locations.

3.4 Physico-Chemical Profile of Dried Mangoes Sold in Chad: Principal Component Analysis (PCA) and Hierarchical Ascending Classification (HAC)

The water content is shown in Table 1. According to Sawadogo-Lingani (1993), it influences the preservation and perishability of foodstuffs, and, especially in mango [8]. A water content higher than 14% favors microbial, enzymatic, and chemical activity, thus reducing the shelf life of organics. In fact, this parameter is influenced by variety, climatic conditions, (rainfall), and drying conditions (drying time and type) [33,34]. Water content obtained were all lower than the 14% reported by Ndangui (2015) as the limit for good preservation, especially in tropical zones where humidity generally reaches 50%. The contents obtained were lower than those obtained by Kameni et al. (2003), which ranged from 16.8% to 22.6% for the Amelie, Zill, Irwin and Locale varieties. This can be due to more advanced fruit

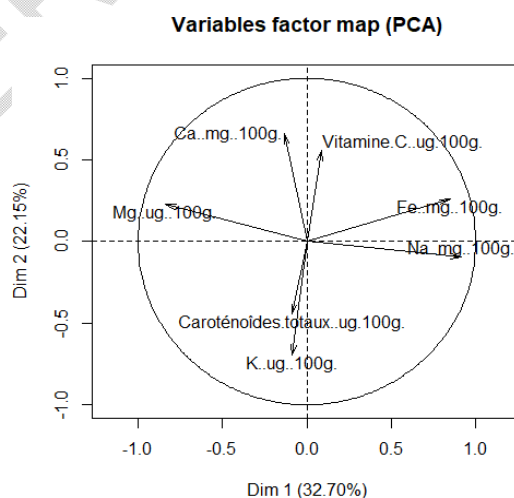


Fig.2.Observations and variables Biplot

Table1. Approximate chemical composition of dried mangoes

Sites	Dba	Bbja	Mdou	Kmr	Bgr	p value
Water content	15.14±0.46 ^b	13.90±0.06 ^a	15.31±0.88 ^b	15.13±0.44 ^b	14.96±0.11 ^b	0.0014**
g/100g						
starchg/100g	111.67±1.85 ^b	110.32±1.33 ^{ab}	110.86±0.47 ^b	108.24±1.49 ^a	112.07±0.54 ^b	0.000994***
Reducing sugars	77.73±0.44 ^a	77.49±0.86 ^a	76.51±0.75 ^a	77.08±0.47 ^a	77.72±1.01 ^a	0.0785
g/100g						
Total carbohydrates	104.33±0.89 ^c	97.20±1.61 ^b	94.91±0.52 ^a	104.28±0.67 ^c	104.72±0.64 ^c	1.06e-13***
g/100g						
Ash(g/ 100g)	2.63±0.07 ^b	2.87±0.02 ^c	2.41±0.04 ^{ab}	2.35±0.05 ^a	2.59±0.25 ^b	1.19e-05***
Proteinsg/100g	3.90±0.10 ^a	3.90±0.06 ^a	3.91±0.02 ^a	3.93±0.05 ^a	3.89±0.09 ^a	0.958
Lipidsg/100g	2.07±0.02 ^c	1.87±0.02 ^{bc}	1.41±0.04 ^a	1.35±0.05 ^a	1.71±0.31 ^b	3.77e-07***
Pecting/100g	94.33±0.89 ^a	97.20±1.61 ^b	94.91±0.52 ^a	94.28±0.67 ^a	94.72±0.64 ^a	0.000484***
Total Fiberg/100g	3.130±0.048 ^b	3.732±0.052 ^c	2.910±0.020 ^a	3.846±0.132 ^{cd}	3.934±0.122 ^d	2.63e-14***
Energykcal/100g	451.572±3.956 ^c	421.364±6.499 ^b	407.972±2.357 ^a	449.010±4.980 ^c	449.426±4.797 ^c	5.98e-13***
Starchg/100g	111.67±1.85 ^b	110.32±1.33 ^{ab}	110.86±0.47 ^b	108.24±1.49 ^a	112.07±0.54 ^b	0.000994***
Reducing sugars	77.73±0.44 ^a	77.49±0.86 ^a	76.51±0.75 ^a	77.08±0.47 ^a	77.72±1.01 ^a	0.0785
g/100g						
Total sugarsg/100g	104.33±0.89 ^c	97.20±1.61 ^b	94.91±0.52 ^a	104.28±0.67 ^c	104.72±0.64 ^c	1.06e-13***
Amylopecting/100	94.334±0.892 ^a	97.198±1.613 ^b	94.906±0.524 ^a	94.282±0.672 ^a	94.722±0.635 ^a	0.000484***
Amyloseg/100g	19.512±0.308 ^c	17.420±0.674 ^b	15.112±0.414 ^a	20.910±0.412 ^d	15.052±0.360 ^a	4.49e-15***
Amylose/ Amylopectin	4.830±0.112 ^b	5.580±0.160 ^c	6.278±0.142 ^d	4.506±0.091 ^a	6.292±0.107 ^d	3.98e-16***

Means ± standard deviations followed by the same letter in the same row indicate that differences are not significant ($P > 0.05$). Dba: Doba; Bbja: Bebedjia; Kmr: Koumra; Mdou: Moundou; Bgr: Bongor

ripening and use of a slow-flow solar dryer, the reduced drying time, cultivation in an area (Cameroon) with high rainfall and humidity, and the variety of mangoes used, which are richer in water as explained by Ndangui [33] and Tambo [34]. Indeed, it has been reported that the use of more mature fruits resulted in dried products with a higher water content [35]. The results obtained by Bélem are in the same range (4.14 to 4.19%) as those of this study [11]. Thus, these results demonstrate that dried fruit, regardless of locality, can be preserved over a very long period of time (more than 6 months).

Ash represents the inorganic part of the material and thus provides information on its mineral content. A content less than 5% is recommended for supplementary feeds [36]. Table 6 shows that the samples obtained significantly ($p = 0.05$) influenced this parameter. Overall, variation was significant ($p = 1.060.05$) between localities. The content varied between 2.35 and 2.87 g/100g. The high content observed in the Bebedja sample is thought to be linked to the variety used in this locality, climatic conditions, particularly rainfall, which is lower and therefore favors the absorption of minerals present in the soil, the nature of the soil and its composition, and the type of drying applied. Mwamba et al. [37] also reported the influence of drying type on ash content, demonstrating the positive impact of oven drying. In fact, solar drying leads to prolonged exposure of the food matrix, resulting in a possible loss of minerals by diffusion with the eliminated water.

The contents obtained are in the same range as those of previous studies [11,37], which ranged from 1.59 to 2.60% and 1.98 to 2.57%, respectively. However, they were lower than those of Rakotonantoandro [38], which ranged from 4.27 to 7.46%. These results show that these dried fruits could be used in the formulation of food supplements. Proteins play many

physiological and structural roles in the body. Their proportions in dried fruits depend on drying conditions (drying time and temperature), variety, climatic conditions, the nature of the soil, and the state of maturity of the plant [11]. Protein content ranged from 3.89 to 3.93 g/100 g. It was not significantly ($p = 0.906$ to 0.05) affected by area of origin. The high protein content in samples collected at Koumra could be explained by the overexpression of protein synthesis genes to the detriment of lipids and carbohydrates in this variety. The negative correlation observed between this parameter and carbohydrates ($r = -0.6322$) and lipids ($r = -0.8477$) confirms this assertion. In addition, drying at temperatures above 60°C by arboriculturists in the four other localities would have resulted in a loss of protein due to the formation of melanoids during the Maillard reaction. The values obtained are five times higher than those of Traore [39] and Mwamba [37]. These results show that dried mangoes from different localities could easily be used to supplement porridges and thus combat protein-energy malnutrition.

Lipid content was not significantly different ($p < 0.05$). It ranged from 1.35 to 2.07 g/100 g. Overall, contents were below the 5-8% recommended by the Codex Stan (2013) standard for the formulation of supplementary feeds. Indeed, many authors ([10,37,39] have reported the non-lipidic food character of fruits. The drop in this parameter in samples from Koumra and Bongor is linked to varietal differences, with advanced ripening in fruit from these localities resulting in a loss of this compound, as well as the type of drying used. Mwamba et al. [37] reported a low lipid content in solar-dried fruit, probably due to oxidation. The values obtained are higher than those of Nabalma [40], Sawadogo-Lingani et al. [8], and Traore [39], which ranged from 0.6% to 1.85%. The use of this fruit therefore requires supplementation with lipid sources.

Table 2. Composition and mineral ratios of dried mangoes

Sites	K(ug/100g)	Mg(ug/100g)	Fe(mg/100g)	Ca(mg/100 g)	Na(mg/100 g)
Db	85.97±4.93 ^a	70.24±0.46 ^c	1.05±0.11 ^a	73.25±0.52 ^{ab}	11.41±0.29 ^a
Bbj	80.92±6.94 ^a	70.59±0.38 ^c	1.50±0.45 ^{ab}	73.35±0.46 ^{ab}	11.50±0.45 ^a
Md	83.21±1.81 ^a	70.06±0.37 ^{bc}	1.18±0.27 ^a	72.66±0.47 ^a	11.50±0.19 ^a
Km	79.15±0.46 ^a	69.53±0.20 ^b	1.26±0.14 ^a	73.49±0.17 ^a	11.55±0.45 ^a
Bg	82.59±0.25 ^a	68.78±0.30 ^a	1.79±0.20 ^b	72.89±0.07 ^{ab}	12.58±0.39 ^b
pvalue	0.112	8.97e-07 ^{***}	0.00213 ^{**}	0.0163	0.00028 ^{***}

Means ± standard deviations followed by the same letter in the same row indicate that differences are not significant ($P > 0.05$). Db : Doba ; Bbj : Bebedja ; Km : Koumra ; Md : Moundou ; Bg : Bongor

Table 3. Vitamin composition of dried mangoes

Sites	Vitamine C (mg/100g)	Total Carotenoids (ug/100g)
Db	69.37±0.58 ^a	265.97±343.34 ^a
Bb	70.05±0.45 ^a	249.25±412.35 ^a
Md	70.04±0.36 ^a	255.44±422.12 ^a
Km	70.15±0.79 ^a	274.15±466.70 ^a
Bg	69.68±0.37 ^a	258.48±432.98 ^a
pvalue	0.161	0.906

Means±standard deviations followed by the same letter in the same row indicate that differences are not significant ($P>0.05$)

Table 4. Contribution of observations to the formation of the different axes

Samples	Contributions of observations (%)			Square cosines of observations			Class
	F1	F2	F3	F1	F2	F3	
Db	23.3482	13.8300	2.1580	0.5272	0.2435	0.0215	1
Bb	6.2927	6.9211	12.3208	0.2136	0.1832	0.1848	1
Km	7.8684	15.3444	55.1842	0.1765	0.2684	0.5469	2
Md	7.4257	45.3802	24.2244	0.1371	0.6531	0.1975	2
Bg	55.0650	18.5243	6.1125	0.7619	0.1998	0.0374	3

Table 5. Contribution of variables to the formation of the different axes

	Contributions of variables (%)			Square cosines of variables			Class
	F1	F2	F3	F1	F2	F3	
Vitamine C	5.7826	2.2975	2.7268	0.5434	0.1683	0.1132	1
Carotenoids	0.1722	11.6803	0.0601	0.0162	0.8558	0.0025	2
Reducing sugars	7.4852	3.8362	0.2027	0.7034	0.2811	0.0084	3
Water content	2.3600	4.8182	0.3313	0.2218	0.3530	0.0138	3
Lipids	1.9031	10.4314	1.3693	0.1788	0.7643	0.0568	3
Starch	6.0477	2.7401	3.4436	0.5683	0.2008	0.1429	3
Amylose	4.6870	2.5487	6.1320	0.4404	0.1867	0.2545	3
Amylopectin	4.6870	2.5487	6.1320	0.4404	0.1867	0.2545	4
Amylose/amylopectin	4.1108	3.4623	6.3003	0.3863	0.2537	0.2615	3
Mg	10.3671	0.1794	0.2091	0.9742	0.0131	0.0087	4
Ca	5.7984	0.2838	10.3986	0.5449	0.0208	0.4316	5
Proteins	4.0991	3.6508	7.6507	0.3852	0.2675	0.3175	3
Ash	0.7467	0.0696	16.9053	0.0702	0.0051	0.7016	3
Na	9.4570	1.1011	0.4067	0.8886	0.0807	0.0169	4
K	5.3357	6.3428	0.7932	0.5014	0.4647	0.0329	6
Fibers	2.5936	8.7608	2.7409	0.2437	0.6419	0.1138	3
carbohydrates	3.3814	8.6470	1.1311	0.3177	0.6336	0.0469	4
Energy	1.3864	7.7840	6.6989	0.1303	0.5703	0.2780	7
Fe	0.2719	10.3553	3.8054	0.0255	0.7587	0.1579	3

Digestible carbohydrates represent the main form of metabolites that can be metabolized by the body for energy production. Its content ranges from 76.51 to 77.73 g/100 g with non-significant variation ($p>0.05$) between different samples. Carbohydrates depend on the proportion of lipids and proteins in the plant (Tambo et al., 2019a,b). The values obtained are higher than those of Nabalma et al. [40,39,37], which were 61.81%, 58.82-63.11% and 68.52% respectively. A variety of

difference associated with inadequate drying conditions would explain these differences. Indeed, Mwamba et al. reported an improvement in total carbohydrate content with oven drying compared to solar drying [37]. The contents obtained by Kameniet al. [41,11] are in the same range as those of this study. The values obtained are broadly in line with the Codex Stan standard, which recommends a carbohydrate content of between 65 and 85% [42].

Table 6. Pearson correlation matrix (r) of the different variables

Variables	VITC	Carotenoids	Reducing sugars	TE	Lipides	Amidon	Amylose	Amylopectine	Amy/amyopMg	Zn	Ca	Protéines	Cendres	Na	K	Fibres	Glucides	Energie	Fe
VITC	1																		
Caroténoides	-0,3418	1																	
Sucres réducteurs	-0,8403	0,5626	1																
TE	0,8988	-0,3881	-0,7534	1															
Lipides	0,0362	0,7456	0,1301	-0,2877	1														
Amidon	0,7365	-0,3864	-0,9295	0,7672	-0,1609	1													
Amylose	-0,3528	0,5809	0,8029	-0,2895	0,2199	-0,7826	1												
Amylopectine	0,3528	-0,5809	-0,8029	0,2895	-0,2199	0,7826	-1,0000	1											
Amylose/amylopectine	-0,3614	0,6307	0,8086	-0,3306	0,3017	-0,7944	0,9961	-0,9961	1										
Mg	-0,7168	0,2494	0,8917	-0,4814	-0,2945	-0,8118	0,7733	-0,7733	0,7387	1									
Ca	-0,8023	0,2787	0,6308	-0,4767	-0,3423	-0,3571	0,2388	-0,2388	0,2119	0,6872	0,7725	1							
Protéines	-0,3626	-0,3102	0,1800	0,0597	-0,8477	0,0281	-0,0365	0,0365	-0,1087	0,5117	0,8551	0,7628	1						
Cendres	0,2501	-0,1768	-0,0675	-0,1224	0,3707	-0,2891	0,1152	-0,1152	0,1514	-0,2051	-0,4748	-0,7603	-0,7549	1					
Na	-0,5729	-0,0948	0,6182	-0,2150	-0,6771	-0,4994	0,4777	-0,4777	0,4133	0,8932	0,9981	0,7464	0,8254	-0,4343	1				
K	-0,7275	0,7227	0,9693	-0,6972	0,3400	-0,8986	0,8668	-0,8668	0,8860	0,7959	0,4250	0,5034	-0,0100	-0,0017	0,4540	1			
Fibres	0,0677	-0,7042	0,0223	0,2669	-0,8290	-0,1482	0,1429	-0,1429	0,0679	0,4252	0,6315	0,0260	0,5263	0,1065	0,6462	-0,1362	1		
Glucides	0,0336	0,6903	-0,0742	-0,2060	0,8829	0,1626	-0,1250	0,1250	-0,0471	-0,4826	-0,7160	-0,1568	-0,6322	0,0048	-0,7244	0,1055	-0,9901	1	
Energie	-0,1600	0,7308	0,0369	-0,2468	0,6882	0,1764	-0,1289	0,1289	-0,0677	-0,3095	-0,4460	0,1966	-0,2922	-0,3612	-0,4691	0,1681	-0,9648	0,9249	1
Fe	-0,4739	0,7201	0,3116	-0,6431	0,7326	-0,1909	-0,0129	0,0129	0,0607	-0,1101	-0,3586	0,2561	-0,3672	-0,1148	-0,3747	0,4010	-0,9047	0,8594	0,8899 1

Les valeurs en gras sont différentes de 0 à un niveau de signification $\alpha=0,05$. VITC: Vitamine C; TE: Teneur en eau; Amy/amyop: Rapport Amylose/Amylopectine.

Fiber represents carbohydrate fraction not digestible by enzymes of the human gastrointestinal tract, and is responsible for intestinal transit speed. Fiber is inversely related to digestible carbohydrates ($r = -2.63 \cdot 10^{-14}$). Levels were significantly ($p = 2.63 \cdot 10^{-14} < 0.05$) affected by production locality, and consequently ranged from 2.910 (Moundou) to 3.94 g /100g (Bongor). The lower content observed in Koumra samples can be explained by the advanced state of maturity due to the degradation of fibers by microorganisms, producing sugars of lower molecular weight. In addition, oxidation of these fibers into organic compounds by intrinsic enzymes as well as by microorganisms would also reflect this lowering [41]. Kameni et al. obtained fiber contents between 0.7 and 1.80% in four varieties of dried mango, which is far lower than what we obtained in this work [41]. Overall, the values obtained are in line with the Codex Stan standard, which recommends less than 5.60% fiber in supplementary foods [42].

Energy density was also assessed, showing that dried mangoes from Bebedja and Moundou had the lowest and highest energy densities, respectively. This parameter ranged from 407.972 ± 2.357 to 451.572 ± 3.956 kcal/100g of DM. The high energy density observed with the Doba and Moundou samples is mainly linked to their high lipid content, as confirmed by the positive correlation ($r = 0.6882$) between these two parameters. There was also a significant influence ($p = 5.98 \cdot 10^{-13} *** < 0.05$) of production area on energy density. The values obtained are on the whole lower than those recommended (720 kcal) by FAO/WHO [36] to cover the energy needs of weaning-age children and thus effectively combat protein-energy malnutrition. These results suggest supplementation with lipid sources such as soy [43]. The values obtained are similar to those of Eucharia et al. (2020), which ranged from 359 to 361.44 kcal.

Reducing sugars are responsible for fruit sweetness and consumer acceptability. The content of this parameter evolves positively with ripening, unlike starch content, as shown by the negative correlation coefficient ($r = -0.9295$) between the two parameters. Indeed, during ripening. Amylases naturally present in fruit degrade starch into amylose residues, releasing low-molecular-weight carbohydrates such as dextrins, fructose, glucose and maltose [11,41]. The evolution of reducing sugar content is associated with those of carotenoids ($r = 0.5626$) but contrary to those of vitamin C ($r = -0.8403$)

and amylopectin ($r = -0.8029$). Reducing sugar content was significantly ($p = 0.001 < 0.05$) affected by powder origin. Samples from Doba had the highest content ($77.73 \pm 0.44b$), while those from Moundou had the lowest ($76.51 \pm 0.75a$). Samples from Doba and Bongor were statistically similar, which could be explained by the use of similar varieties in both localities, as well as drying at almost similar stages of ripeness [11]. The values obtained are lower than the 39.30% and 48.95% reported by Bélem et al. [11] on the Amelie and Brooks varieties from Burkina Faso respectively. Similarly, 75.06% obtained by Djantou [44] on Kent was higher. These differences are linked to the drying conditions, the varieties studied and the climatic conditions in the growing areas. Indeed, drying at high temperatures would lead to complexation of simple sugars at their aldehydic or ketonic functions with amine groups, thus reducing their content [11]. On the other hand, these results are similar to those obtained by Kameni [41] which ranged from 10.80 to 24.70%. The results obtained show that the various dried mangoes could be used as a natural sweetening agent in supplementary foods and thus limit the dangers associated with refined sugars.

Starch, amylose and amylopectin contents were significantly ($p < 0.05$) affected by dried mango sources. They ranged respectively from 108.2 to 112.07 g/100g starch. The difference in varieties with other localities, the less advanced state of ripening, the storage in a dry environment with no activity, the high starch content of Bongor powders can be explained by the consequent microbial presence, the absence of microorganisms with amylase activity and the drying conditions. Kameni et al also reported a positive relationship between low ripening and starch content in mangoes [41]. The starch contents obtained are higher than those reported by Djantou [44] in his study, which ranged from 1.60 to 1.69%. This can be explained by the higher water content in this author's matrices which has a diluting effect on nutrients. Indeed, many studies have reported an increase in nutrient concentration in dried mangoes with decreasing water activity [45,46]. The opposite trend was observed between amylose and amylopectin in all samples as demonstrated by the negative correlation coefficient ($r = -1.000$) between the two parameters. Indeed, Tambo et al., 2019a.b and Dongmo et al., 2020 it have been reported such observations in several studies [34,47,48]. The disparity observed in the composition of these two macronutrients is

thought to be directly related to the nature of starch (degree of branching), the mango variety, storage conditions and the pretreatments applied. The amylose/amylopectin ratio is an indicator of the rheological properties of a flour or powder and its ability to be used in formulations [49]. It is highly dependent on amylose composition (a positive correlation of 0.9961 between the two parameters). It follows from Table 2 that this parameter was significantly ($p < 0.05$) affected by the source of dried mangoes. The Amylose/Amylopectin ratio ranged from 4.506 (Koumra) to 6.278 (Moundou). These results demonstrate the high heat-treatment instability of dried mangoes from Moundou. The higher the value, the greater the tendency of the matrix to retrograde following heat treatment, making the formulations unstable. The use of dried mangoes from Moundou in the formulation of supplementary feeds should therefore be carried out at the end of cooking, to avoid thick indigestible porridges [48].

Mineral malnutrition or hidden hunger is the most widespread form of malnutrition today. It affects more than one child in three and is therefore a major health problem for which solutions are constantly being sought [36]. Minerals play many physiological roles in the body, including cofactor of metabolic reactions (Ca, Fe, K, Na, Mg), transmission of nerve impulses, bone solidification, second messenger (calcium), maintenance of osmotic pressure (sodium and potassium), muscle contraction (calcium), bone rigidity and bone formation (phosphorus, calcium and magnesium), energy production, heart function, formation and function of many proteins such as hemoglobin involved in oxygen transport [50-52]. Results from Table 4 showed that all minerals were significantly affected by locality of origin with ($p < 0.05$) respectively for calcium, iron, magnesium, sodium, potassium and copper. Levels varied from 79.15 ± 0.46 (Koumra) to 85.97 ± 4.93 $\mu\text{g}/100\text{g}$ (Doba), from 68.78 ± 0.30 (Bongor) to 70.59 ± 0.38 $\mu\text{g}/100\text{g}$ (Bebedja), from 1.05 ± 0.11 (Doba) to 1.79 ± 0.20 $\text{mg}/100\text{g}$ (Bongor), from 72.66 ± 0.47 (Moundou) to 73.49 ± 0.17 $\text{mg}/100\text{g}$ (Koumra), from 11.41 ± 0.29 (Doba) to 11.55 ± 0.45 $\text{mg}/100\text{g}$ (Koumra) for potassium, magnesium, iron, calcium and sodium respectively. Overall, dried mangoes from Doba, Moundou and Bebedja had the highest mineral content, unlike those from Koumra and Bongor. The differences noted between the different fruits are thought to be the result of variations in the varieties used. The drying method used (particularly solar drying)

which leads to greater mineral loss; the high drying temperature, which could facilitate increased diffusion of ions. Climatic conditions, storage conditions and fruit maturity [8, 37, 41, 46]. Calcium levels were 20 times higher than those of Djantou [44], similar for potassium but lower for sodium and magnesium. Iron content was lower than Sawadogo et al. [8] and Mwamba et al. [37]. at 6 and 7 $\text{mg}/100\text{g}$ respectively. Comparison with recommended daily values shows that all samples (E1 DOBA and E2 BGA) are within the recommended range for calcium (1000 mg). These results suggest a high consumption of dried mangoes. For iron, magnesium and sodium they were all below the recommended daily requirements of 8-18 $\text{mg}/100\text{g}$, 127-1500 $\text{mg}/100\text{g}$ and 8-12 mg respectively. This suggests a need for supplementation with other sources of these minerals in all samples [43, 49].

Vitamins A and C play numerous physiological roles in the body. They prevent the onset of diseases such as cancer through their antioxidant potential, protect against night blindness and strengthen the immune system and intellect [25]. Carotenoid content was significantly ($p > 0.05$) unaffected by collection area. Amounts varied from 249.25 ± 412.35 $\mu\text{g}/100\text{g}$ DM for Bebedja to 274.15 ± 466.70 $\mu\text{g}/100\text{g}$ DM for Koumra. The difference observed can be explained by the drying conditions (time-temperature pair), the degree of sunshine in the locality, the ripeness of the mango, the type of dryer used (solar or electric) and the variety of mango [8, 37]. Indeed, the low values obtained with certain samples can be explained by the use of solar drying which would lengthen exposure resulting in oxidation of carotenoids and loss through evaporation [37]. Similarly, mangoes from this locality would be at a less advanced stage of ripening, thus reducing carotenoid content. Indeed, Some et al. (2014) reported an improvement in carotenoid content as the ripening stage advanced (39 to 80%). The values obtained would enable daily vitamin A requirements to be covered by more than 80%. This suggests the use of dried mangoes in the formulation of substitute foods for children and the elderly. The values obtained are higher than those of Savadogo et al. [53] who worked on Kent, Keitt and Springfield varieties from three towns in Burkina Faso (Orodara, Banfora and Bobo-Dioulasso).

As far as vitamin C is concerned, an increase is observed with the reduction of carotenoids in the

various samples, as demonstrated by the negative correlation ($r = -0.3418$) obtained between the two parameters. Indeed, studies reported that younger mango fruits had higher vitamin C contents, as these degrade into other compounds such as furfural as maturity advances [8,54]. He also found that there was no significant difference ($p = 0.161 > 0.05$) between all the other samples. Results ranged from 69.37 ± 0.58 mg/100 g of dried mangoes for Doba to 70.15 ± 0.79 mg/100 g of dried mangoes for Koumra. These results suggest that traders in Doba use less mature fruit, unlike those in other localities. Similarly, the varieties used in this locality are genetically more inclined to synthesize vitamin C. In addition, the richness in reducing sugars in samples from localities such as Koumra and Moundou could induce Maillard reactions during drying at high temperatures, leading to vitamin C degradation [37]. The results obtained are in the same range as those of Bélem et al. [11], which ranged from 116.20 mg/100 g DM for the commercially mature Amelie variety to 205.77 mg/100 g DM for the immature Amelie variety. Adeyemi et al. obtained lower results than this work (between 77.57 and 53.24 mg/100 g DM in melon-based fruit juice). These results demonstrate the benefits of using melon powders in high-fat dishes that are susceptible to oxidation, as well as recommending them to people suffering from obesity and cardiovascular disease [55].

Principal Component Analysis (PCA) was applied to investigate the association between the physico-chemical properties which are variables and the different samples representing observations. Hierarchical Ascending Classification (HAC), which permit to group observations according to their physico-chemical affinities, gave us 3 classes (Table 4). Small group correlations show that samples from Db, Bb, Kmr, Mdou and E5Bgr form 1, 1, 2, 2 and 3 classes respectively. Kameni et al. also reported a certain similarity in the physicochemical composition of dried mango fruits from different varieties at different commercial maturity [35,41]. Table 4 also shows that observations Db, Bb and Bgr form the F1 axis, Mdou the F2 axis and Kmr the F3 axis. These results suggest that the variables or physico-chemical properties responsible for the formation of these axes are not significantly different.

The contribution of the variables to the formation of the different axes is presented in Table 5, and shows that vitamin C, reducing sugars, starch,

amylose, amylopectin, amylose/amylopectin ratio, ions such as Mg, Ca, Na, K and proteins are correlated with the Db, Bb and Bgr samples and contribute to the formation of the F1 axis. In fact, samples from Bongor showed the best nutrient contents, surely linked to the best production and storage conditions. Carotenoid, water, lipid, fiber, carbohydrate and iron contents, as well as energy density, are responsible for the formation of the F2 axis. Indeed, observation E4Mdou is rich in carotenoids and water due to its advanced maturity. Ash and the Ca/Mg ratio are responsible for the formation of the F3 axis. Hierarchical Ascending Classification (HAC) of physico-chemical parameters revealed the existence of 7 classes, enabling variables to be correlated with each other.

FIG. 2 shows the Biplot correlation between observations and variables. It can be seen from these figures that observations E1Db and E2Bb are very rich in starch, amylopectin, vitamin C and water content. Indeed, many authors have reported high water content in low-maturity samples [37,55]. In addition, starch, vitamin C and amylopectin content decrease with increasing maturity. These observations suggest that arboriculturists in Doba and Bébedjia harvest and process fruit at an early stage of ripening. Samples from Koumra and Moundou are essentially rich in carotenoids, iron, lipids, carbohydrates and energy. The high lipid and carbohydrate content of these samples is responsible for their high energy density. The advanced maturity of these samples before processing is responsible for their high carotenoid contents, as demonstrated by Bélem et al. [11]. Iron richness is linked to transport by the abundant carotenoids in these matrices. A positive correlation ($r = 0.7201$) between these two parameters confirms these observations. The third group consists of the Bongor sample, which is associated with proteins, fibers and ions such as Ca and Na. The overexpression of genes responsible for the synthesis of proteins and other ions in the variety used in the Bongor locality are at the origin of these results.

Vitamin C and carotenoids form classes 1 and 2 respectively. Plants have the capacity to synthesize these powerful antioxidants and plant colorants in their free state within the plant.

The second class is made up of reducing sugars, water content, lipids, starch, amylose, amylose/amylopectin ratio, iron, proteins, ash

and fiber. In this class, we note a negative correlation between water content and the elements amylose ($r = -0.2895$), reducing sugars ($r = -0.7534$; $p < 0.05$), iron ($r = -0.6431$; $p < 0.05$). Indeed, Bélem et al. (2017) reported that a decrease in water content increased the concentration of the various nutrients in the matrix and thus their contents. The negative correlation ($r = -0.9295$; $p < 0.05$) observed between reducing sugar and starch content is linked to starch hydrolysis with the formation of simple sugars by the plant's intrinsic enzymes (amylases) during the ripening process. An improvement in the quantity of reducing sugars with amylose content ($r = 0.8029$; $p < 0.05$) would be linked to the low crystallinity of this molecule, unlike amylopectin, which makes it very easily digestible by enzymes (Kenfack et al., 2021). A positive correlation was also observed between amylose and lipids, which is contrary to the work of many authors [43, 35, 47, 56]. The positive association ($r = 0.5263$; $p < 0.05$) between proteins and fibers is linked to the existence of proteins in the form of glycoproteins on the surface of certain membranes [48]. Proteins were also positively associated with Iron ($r = -0.3672$). Protein synthesis requires the action of numerous enzymes that use metal ions such as Calcium and Magnesium as cofactors for their activities. The involvement of ions in enzyme activity is also concentration-dependent [57].

The low iron content in the various samples would thus explain the negative correlation with protein content. Moreover, enzymes being proteins require the presence of ions for their enzymatic activities; they are thus called metalloenzymes [58]. The amylose/amylopectin ratio is positively related to amylose content ($p < 0.05$). Dongmo et al [48] also reported the same observations.

The fourth class is formed by carbohydrates, amylopectin, Magnesium, net Sodium. In fact, the positive improvement in amylopectin content with carbohydrates can be explained by the fact that amylopectin is one of the digestible carbohydrates whose synthesis is generally greater in cereals, fruit and vegetables than in amylose. Due to their high electroneutrality, carbohydrates are incapable of forming electrostatic interactions, hence their negative association with Sodium and Magnesium ($p < 0.05$).

Classes 5, 6 and 7 are respectively formed by Calcium, Potassium and energy value. Ions can

exist in the plant either in a complexed state with other elements (phytates, oxalates...), or in a free state. In the free state, they are more available and therefore easily assimilated. The formation of classes 5 and 6 thus demonstrates good availability of these two ions in dried mango fruit from the five localities.

4. CONCLUSION

The aim of this study was to assess the influence of collection areas on the physico-chemical properties of dried mango fruit. It was found that dried fruit from all localities kept well, as their water content was within the norm. However, several drying methods from different collection areas didn't significantly modify some physico-chemical properties of dried mangoes fruits. Therefore, dried mangoes fruits from these areas could have good nutritional value and could be used as food supplements.

REFERENCES

1. SNV (Netherlands Development Organisation) Project to support the marketing of dried mangoes and processed cashew nuts. Memorandum of Understanding between the United Nations Office for Project Services and the Government of Burkina Faso for the implementation of the Category 2 Project of the Enhanced Integrated Framework. 2014;123.
2. Vannière H, Didier C, Rey JY, Diallo T, Kéita M, Sangaré S. M. Lamangou en Afrique de l'ouest francophone: Les systèmes de production et les itinéraires techniques Fruits. 2004;59:383–398. Available: <https://doi.org/10.1051/fruits:2005001>
3. Siddiq M, Sogi DS, Roidoung S. Handbook of Mango Fruit. Handb. Mango Fruit; 2017. Available: <https://doi.org/10.1002/9781119014362>
4. Kanté-Traore H, Dufrechou Mr, Le Meur Lay D, Lancon-Verdier V, Dicko MH, Sawadogo-Lingani H. Physico-chemical and biochemical characteristics of 14 poorly popularized mango varieties from Burkina Faso. for better valuation. Sciences and Technology-Applied Natural Sciences. 2020;39(2):2021-245.

5. Aboubakar D, Sorto. Mr, Mbayabe L, Woin N, Bourou S, Gandebe M. Marketing of fruits in the savannahs of Central Africa. Communication at the conference. African savannahs in development: Innovating to last. Garoua. Cameroon. Communication summary book. April 20-24, 2009;58.
6. Guira Mr, Zongo JD. Study of the distribution of varieties grown in mango orchards in western Burkina Faso. Science and technology. Natural Sciences and Agronomy series. 2006;28(1 and 2):63-72.
7. ITC (International Trade Center), Mango. Market News Service (MNS). MNS Bulletin. ECOWAS TEN. 2011;26.
8. Sawadogo-Lingani H, Traore SA. Chemical composition and nutritional value of Amélie mango (*Mangifera indica* L.) from Burkina Faso. Journal of Sciences. 2002;2(1):35-39.
9. Djioa T. Improvement in the conservation of 4th range mangoes by application of technical treatments and use of conservation under modified atmosphere. 2010;6.
10. Kouassi OA. National review to identify non-food development initiatives for mango in Ivory Coast. 2012 No. 12/COLEACP PAEPARD-01/BNA-12. September-December 2012;43.
11. Bélem A, Tapsoba F, Touloumé S, Songre-Ouattara L, Zongo C, Savadogo A. Study of the organoleptic quality of three varieties of dried mangos: Amélie. Lippens and Brooks during storage by enzymatic tanning technique of peroxidases (POD) and the polyphenol oxidases (PPO). Revue Scientifique de Technologie et Synthèse. 2017;34:38-47.
12. Djantou NEB, Mbofung CMF, Scher J, Kameni A. Influence of the production process of mango powder (*Mangifera indica*) on some chemical and nutritional characteristics. 2004 AGORALE meetings. Nantes. France; Nov/Dec 2004.
13. Kim D, Jeong SW, Lee CY. Antioxidant capacity of phenolic phyto-chemicals from various cultivars of plums. Food Chemistry. 2003;81:321-32.
14. Chiou A, Karathanos VT, Mylona A, Salta FN, Preventi F, Andrikopoulos NK. Currants (*Vitis vinifera* L.) content of simple phenolics and antioxidant activity. Food Chemistry. 2007;102:516-522.
15. Lee SK, Kader AA. Preharvest and postharvest factors influencing Vitamin C content of horticultural crops. Postharvest Biology and Technology. 2000;20:207-220.
16. Vasquez-Cañedo AL, Sruamsiri P, Carle R, Neidhart S. Accumulation of all-trans carotene and its 9-cis and 13-cis stereoisomers during postharvest ripening of nine Thai mango cultivars. Journal of Agricultural and Food Chemistry. 2005;53:4827-4835.
17. Veda S, Platel K, Srinivasan K. Varietal Differences in the Bio accessibility of β -Carotene from Mango (*Mangifera indica*) and Papaya (*Carica papaya*) Fruits. Journal of Agricultural and Food Chemistry. 2007;55:7931-7935.
18. Manthey JA, Perkins-Weazie P. Influences of harvest date and location on the level of β -Carotene. Ascorbic Acid. Total Phenols. The *In vitro* antioxidant capacity. And phenolic profiles of five commercial varieties of Mango (*Mangifera indica* L.). Journal of Agricultural and Food Chemistry. 2009;57:10825-10830. DOI:10.1021/jf902606h
19. Robles-Sánchez RM, Rojas-Graü MA, Odriozola-Serrano I, González-Aguilera GA, Martín-Belloso O. Effect of minimal processing on bioactive compounds and antioxidant activity of fresh-cut 'Kent' mango (*Mangifera indica* L.). Postharvest Biology and Technology. 2009;51:384-390.
20. Ma X, Wu H, Liu L, Yao Q, Wang S, Zhan R, et al. Polyphenolic compounds and antioxidant properties in mango fruits. Scientia Horticulturae. 2011;129:102-110.
21. Liu F, Fu XSF, Bi XF, Chen F, Liao XJ, Hu X, et al. Physicochemical and antioxidant properties of four mango (*Mangifera indica* L.) cultivars in China. Food Chemistry. 2013;138:396-405.
22. Ara R, Motalab M, Uddin MN, Fakhruddin ANM, Saha BK. Nutritional evaluation of different mango varieties available in Bangladesh. International Food Research Journal. 2014;21(6):2169-2174. Available: <http://www.ifrj.upm.edu.my>
23. Kothalawala SG, Jayasinghe JMJK. Nutritional evaluation of different mango varieties available in Sri Lanka. International Journal of Advanced Engineering Research and Science (IJAERS). 2017;4(7):128-131. ISSN:2349-6495(P)2456-1908(O) Available: <https://dx.doi.org/10.22161/ijaers.4.7.20>

24. WHO, Global prevalence of vitamin A deficiency. Micronutrient deficiency information system. MDIS working paper N° 2. World Health Organisation. Geneva; 1995.
25. Sibetcheu D, NanKap M, Kollo B. Vitamin A deficiency in Cameroon; 1999.
26. Lauricella M, Emanuele S, Calvaruso G, Giuliano M, D'Anneo A; Multifaceted health benefits of *Mangifera indica* L. (Mango): The inestimable value of orchards recently planted in Sicilian Rural areas. *Nutrients*. 2017;20(9):525.
DOI:10.3390/nu9050525
27. Micha R, Penalvo JL, Cudhea F, Imamura F, Rehm CD, Mozaffarian D. Association between dietary factors and mortality from heart diseases. Stroke and type 2 diabetes in the United States. 2017 *JAMA*;317(9):912-924.
DOI:10.1001/jama.2017.0947
28. Martin M, He Q. Mango bioactive compounds and related nutraceutical properties-A review. *Food Rev. Int.* 2009;25:346–370.
Available: <https://doi.org/10.1080/87559120903153524>
29. Tharanathan RN, Yashoda HM, Prabha TN. Mango (*Mangifera indica* L.). The king of fruits- An overview. *Food Reviews International*; 2006.
Available: <https://doi.org/10.1080/87559120600574493>
30. AOAC (Association of Official Analytical Chemists), Official methods of analysis (15th edition). Washington DC. USA. 1990;808-835.
31. Fischer E, Stein E. DNS colorimetric determination of available carbohydrates in foods. *Biochemical Preparation*. 1961;8(1):30 – 37.
32. Chrastyl J. Improved colorimetric determination of amylose in starch or flour. *Carbohydrate Research*. 1987;159:154-158.
33. Ndangui C. Production and characterization of sweet potato flour (*Ipomoea batatas* Lam): Optimization of bread-making technology. Doctoral thesis in food processes and biotechnologies defended at the University of Lorraine and the University of Marien Ngouabi. Brazzaville. Congo. 2015;135.
34. Tambo TS, Klang JM, Ndomou HSC, Kohole FHA, Womeni HM. Application of amylase-rich flour of corn and sweet potato to the reduction of consistency of cassava and corn gruels. *Journal of Food Processing and Preservation*. 2019b;43(9):e14058.
Available: <https://doi.org/10.1111/jfpp.14058>
35. Kameni A, Mbofung CM, Ngnamtam Z, Doassem J, Hamadou L. Ability to dry some varieties of mango grown in Cameroon: Amélie. Zill. Irwin and Horé Wandou. Proceedings of the conference. Garoua. Cameroon; May 27-31 2002.
36. FAO/WHO, Joint FAO/WHO Food Standards Program. Report of the twenty-seventh sessions of the Codex Committee on Nutrition and Foods for Special Dietary Uses. ALINOM 06/29/26. 2006;105.
37. Mwambal, Tshimenga K, Mbabu D, Conas M, Mputu Kanyinda JN. Séchage des mangues par étuvage et au soleil. *International Journal of Innovation and Scientific Research*. 2018;37(2):216-222.
38. Rakotonantoandro LV. Valorization of the mango: Pastes and pickles. Final Dissertation in Agricultural and Food Industries. 2010;117.
39. Traore S. Stabilization and conservation of mango pulp for the production of fermentation vinegar. End of Study dissertation for obtaining the Professional License in Biological Engineering. 2013;56.
40. Nabalma DF. Thesis with a view to obtaining a higher education diploma specialized in agri-food industries. 1995;45.
41. Kameni A, Mbofung CM, Ngatam Z, Doassem J, Hamadou L. Ability to dry the fruits of some varieties of mango trees grown in Cameroon. *Fruits*; 2003.
42. Codex Alimentarius, Guidelines for the development of complementary food preparations for older infants and young children (CAC/GL08-1991.rev.2013); 2013.
43. Klang JM, Tambo TS, Doungmo FA, Tsopbeng TAB, Womeni HM. Application of germinated corn flour on the reduction of flow velocities of the gruels made from corn. Soybean. *Moringa oleifera* leaf powder and cassava. *Journal of Food Processing and Technology*. 2019a;10(7):800.
DOI:10.4172/2157-7110.1000800
44. Djantou EB. Optimization of the grinding of dried mangoes (*Mangifera indica* var Kent): Influence on the physicochemical and functional properties of the powders

- obtained. Vandoeuvre-les-Nancy. INPL. 2006;151 pp.
45. Jiokap NY, Nuadge GB, Raoul-Wack AL, Giroux F. Dehydration - Impregnation by immersion of mango slices (*Mangifera indica*): Influence of the temperature and the concentration of the solution on the kinetics of certain constituent elements of the fruit. *Fruits*. 2001;56:163-177.
 46. Saliha E, Sahin MG, Selahattin. The effects of cutting and drying medium on the Vitamin C content rosehip during drying. *Journal of Food Engineering*. 2005;68(4):513-518.
 47. Tambo TS, Klang JM, Ndomou HSC, Teboukeu BG, Womeni HM. Characterisation of corn. Cassava and commercial flours: Use of amylases rich flours of germinated corn and sweet potato in the reduction of the consistency of the gruels made from these flours-influence on the nutritional and energy value. *Food Science and Nutrition*. 2019;7(4):1190–1206.
Available: <https://doi.org/10.1002/fsn3.902>
 48. Dongmo H, Tambo TS, Teboukeu BG, Mboukap NA, Fotso SB, Djuidje TMC, Klang JM. Effect of process and variety on physico-chemical and rheological properties of two new corn flour varieties (ATP and Kassai). *Journal of Agriculture and Food Research*. 2020;2:10075.
Available: <https://doi.org/10.1016/j.jafr.2020.100075>
 49. Klang JM, Tambo TS, Nguemguo KLG, Teboukeu BG, Ndomou HSC, Kohole FHA, Womeni HM. Effect of bleaching and variety on the Physico-chemical. Functional and rheological properties of three new Irish potatoes (Cipira, Pamela and Dosa) flours grown in the locality of Dschang (West region of Cameroon). *Heliyon*. 2019b;5(12):e02982.
Available: <https://doi.org/10.1016/j.heliyon.2019.e02982>
 50. FAO, Improving nutrition through home gardening. A training package for preparing field workers in Africa. FAO Rome; 2001
Available: <http://faostat3.fao.org/browse/FB/CC/E>. Accessed 2015 July 12
 51. Badham J, Zimmermann B, Kramer K. The guide to nutritional Anemia. Switzerland: Sight and Life. 2007;56-64.
 52. Ogbonnaya A, Ketiku C, Mojekwu J, Energy. Iron and zinc densities of commonly consumed traditional complementary foods in Nigeria. *British Journal of Applied Science and Technology*. 2012;2(6):48–57.
 53. Savadogo B, Bationo F, Some EN, Ouattara-Songre L, Parkouda C. Détermination de la teneur en β -carotène de trois variétés de mangues consommées au Burkina Faso: Kent, Keitt et Springfield. *International Journal of Innovation and Applied Studies*. 2023;39(1):184-192.
 54. Somé TI, Sakira AK, Tamimi ED. Determination of β -carotene by High Performance Liquid Chromatography in Six Varieties of Mango (*Mangifera indica* L.) from Western Region of Burkina Faso. *The American Journal of Food and Nutrition*. 2014;2(6):95-99.
 55. Adeyemi GF, Chinyelu IN, Ebele I, Olalekan OA, Babatunde A. Effect of preservatives on the physicochemical properties of watermelon and soursop fruit blend. *Direct Research Journal of Agriculture and Food Science*. 2017;5(10):333-337.
Available: <https://doi.org/10.26765/DRJAFS.2017.8912>
 56. Kenfack PSR, Mouamfon M, Tambo TS, Nguemguo JR, Matenchi PY. Effects of three culinary treatments on the physico-chemical, functional and antioxidant properties of wild yam (*Dioscorea praehensilis* Benth.) at East Cameroon. *International Journal of Biological and Chemical Sciences*. 2021;15(6):2665-2684.
Available: <https://dx.doi.org/10.4314/ijbcs.v15i6.32>
 57. Tambo TS, Adebo OA, Ndinkeh TD, Obilana OA, Kohole FHA, Kenfack OJ, et al. Optimization using Response Surface Methodology of proteolytic activity of two maize varieties (Atp-Y and Coca-sr): Amino acids composition. Physico-chemical and functional properties of optimal sample. *Frontiers in Nutrition*. 2023;10:1163915.
Available: <https://doi.org/10.3389/fnut.2023.1163915>
 58. Tambo TS, Klang JM, Ndomou HSG, Teboukeu BG, Kohole FHA, Womeni HM. Characterization of crude extracts amylase flours of corn malt (Kassai and ATP varieties) and sweet potato (Local and

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